THE OBJECTIVES OF THIS WORK

1) V. parahaemolyticus, like other non-cholera Vibrio species, contaminates most marine animals in coastal waters, and causes frequently food poisoning (associated gastroenteritis), wound and soft tissue infections, septicemia, and other infections. Among several thousand of infected persons worlwide per year, there are over 10% cases with severe diseases that may end in death when immunocompromized persons are infected. To get more insight into the multidrug resistance (MDR) mechanism of this microbe, particularly VmrA efflux pump and its function, is one of the objectives:

2) To perform QSAR (Quantitative Structure-Activity Relationship) & chemometric study of structurally unrelated substrates of the VmrA. as extruded by E. col/strains: KAM32 and KAM32/oVCJ6 (with VmrA):

3) To develop QSAR and SAR means to predict to which drugs and xenobiotics V. parahaemolyticus will be sensitive or resistant due to its efflux ability by means of the VmrA pump.

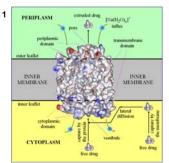
THE VmrA PUMP, ITS MULTIDRUG RESISTANCE (MDR) EFFLUX MECHANISM, AND ITS SUBSTRATES

Vibrio cells

7

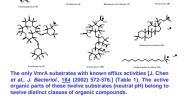
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OXO



Structure of the Nar/multidrug transporter VmrA and drug efflux a Gram-negative cell of V. parahaemolyticus A0334 [M. M. C. Ferreira, R. Kiralj, unpublished], as modeled from primary structure [J. Chon et al., J. Bacteriol, 134 (2002) 575-576.] Influx of Nar in salty medium causes complex formation of Nar with Asp and Glu in transmembrane, cytoplasmic, and periplasmic domains of VmrA. This provekes allosteric changes in the VmrA, and opening the periplasmic window, pore. Drug molecules may be are captured by the inner leafted of the inner membrane and by lateral diffusion are brought to the vestibule. They may be captured directly by the topolasmic domain and brought to the vestibut through which they enter into the central cavity where they accumulate before being extruded from the pump. UmrA is a secondary active transport that uses electrochemical potential of Nar across the membrane as its energy source. It is a defense mechanism of V. parahaemolyticus against several structurally unrelated drugs and xenobiotics.





		0						
o.	CSD source ^a	Formula ^b	pMIC(pVCJ6)*	pMIC(KAM)d	pMIC∆°			
	ASUMEG	C14H13N3	3.242	5.961	2.719			
	DURDAV01	[C ₂₄ H ₂₀ P]*	3.468	4.672	1.204			
f		[C ₁₄ H ₁₄ N ₁] ⁺	3.909	5.114	1.205			
	ETHIDB	[C ₂₁ H ₂₆ N ₃]*	4.392	4.994	0.602			
	CLMPCL02	[C11H12Cl2N2O5]*	5.810	5.810	0			
	XAYGEJ	C16H18FN3O3	7.027	7.027	0			
	QIMMEE	[C ₂₃ H ₃₁ N ₂ O ₃]*	4.777	4.777	0			
	TETCYH10	[C ₂₂ H ₂₅ N ₁ O ₈]*	5.949	5.949	0			
	NAVTEJ	[C ₁₇ H ₆₀ O ₁₃ N]*	5.264	5.264	0			
0	STOSEH10	C ₂₁ H ₄₁ O ₁₂ N ₇]*	5.464	5.464	0			
1	GOLWIN	[C34H3004]	5.318	5.318	0			
2	SATLUU	[C ₁₂ H ₂₅ O ₄ S] ⁻	6.461	6.461	0			
SD codes for the structures retrieved from the CSD database. Complete or partial structures were								
		modeling, bFormula						
ionic/protonated (+), anionic (-) or zwitterionic (±) state as applied in molecular modeling,								
flux activity pMIC of E. coli strain KAM32/pVCI6. 4Efflux activity pMIC of E. coli strain								
M32. 'Difference between the two efflux activities. 'The simplest drug that was not found in the								
D, and was modeled by Titan program.								

Table 1. Experimental data for drugs 1 - 12

Table 5. Prediction of the MDR character of V.

(PCR) character prediction

No. pMICA pMICA MDR

0.951 resistant

0.512

0.313 cancitina

(PLS)

1.687 1.720 resistant

1.710 1.821 resistant

1.240 1 211

0.454

0.340

3

8

Appropriate structures from the Cambridge Structural Database Appropriate structures from the Cambridge Structural Database (CSD) were retrieved and modified, then optimized at PMA semi-empirical level. The activities are pMIC = -log(MIC/moli⁻), where MIC is Minimal Inhibitory Concentration of the drugs as extruded by two strains of E. coli: KAM32 strain without VmrA, and KAM32b/VC36 with. VmrA (VmrA from V. parahaemolyticus AQ334), The pMICA is a measure of the MDR effect of VmrA, defined as pMICA_a abspMIC(VC36) - pMIC(KAM32).

> Correct No. pMICA pMICA

. 22

18 0.820 0.783 resistant

21

to 1-30 by

(PLS) (PCR) 1.137 1.043

0.614 0.643

0.000 0.074

0.635 0.583

0.451 0.482 sensitive

MDR Correc

character prediction

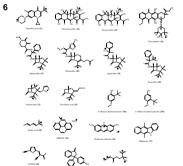
resistant

resistant

MOLECULAR DESCRIPTORS

Min	Symbol	Definition	R[KAM]*	81.01/10				
			- 4 4					
8	ly tr	2nd principal moment of inertia	-0.262	-0.621				
10	FF	3rd principal moment of intertia log(Dip+1), Dip is molecular dipole moment.	-0.216	0.721				
		(Titan)	0.404					
14	Nh	No. hydrophobic carbon atoms	-0.601	-0.200				
15	Na	No. aromatic carbon atoms	-0.469	-0.645				
17	Np	No. polar (not hydrophobic) atoms (non-H)	0.203	0.517				
26	wh	No. ring atoms (non-81) Nh/Nt, number fraction of hydrophobic atoms; Nt	-0.570	-0.410				
-	wn	is total No. non-H atoms	-0.209	-9.726				
28	wa	Na/Nt, mamber fraction of aromatic atoms	+0.387	-0,754				
29	wb	Nb/Nt, number fraction of hydrogen bonding non-	0.562	0.579				
		H atoms; Nb is No. HB donors/acceptors						
30 31	wp wl	Np/Nt, number fraction of polar atoms NUNt, number fraction of planes atoms; NI is No.	0.566	0.726				
21	w1	non-H atoms in all planar fragments	10.1	4.000				
-40	WON	NONNI, number fraction of O/N atoms; NON is	0.454	0.558				
		No. oxygen and nitrogen atoms						
45	Bat	BNt, No. bonds per atom; B is No. bonds (non-	-0.415	-0.603				
		10						
46	WT	Nr/Nt, number fraction of ring atoms	-0.425	-0.713				
50	DM E4	molecular dipole moment (E4 method, MOPAC) average polarizability (E4 method, MOPAC)	0.694	0.788				
55	BT	β hyperpolarizability (E4 method, MOPAC) β hyperpolarizability along the dipole moment	-0.663	-0.456				
		(E4 method, MOPAC)						
59	GM	absolute average y hyperpolarizability (E4	-0.108	-0.508				
		method, MOPAC)						
63	Wn3	W(Nt)3, normalized Wiener index; W is H-	0.510	0.221				
64	BOMO-1	depleted Wiener index energy of HOMO-1 orbital	0.584	0.375				
65	HOMO	energy of HOMO orbital	0.585	0.376				
68	Q-	the most negative ESP atomic charge (non-H)	-0.672	-0.675				
69	Q+	the most positive ESP atomic charge (non-H)	0.426	0.541				
74	Qdf	(Q+) - (Q-), the largest ESP charge difference	-0.539	-0.636				
76	%ikcpk	hcpk/S, surface area fraction of hydrophobic C/H	-0.508	-0.508				
		atoms; hcpk and Scpk are CPK surface areas of hydrophobic C/H atoms and molecule,						
78	Mrefa	respectively MirelN, molecular refractivity per atom: Mirel is molecular refractivity (ClogP method, CliEM3Dr, N is No, all atoms	-0.259	0.696				
79	Enbt	(HOMO-1)/N, HOMO-1 arbital energy per atom	0.233	0.538				
80	Eab	(HUMO)/N, HOMO orbital energy per atom	0.232	0.529				
83	Enb-IIt	[(IOMO-1)/(IOMO)/N. livotier orbital energy	0.232	0.534				
81	Enb-11	sum (HOMO-1)) (HOMO) per atom [(HOMO-1)) (LUMO)]/N, frontier orbital energy	0.328	0.501				
61	ICARD-111	sun (HOMO-1)+(LUMO) per atom	0.319	0.501				
87	DMn	DM/N, DM per atom	0.819	0.788				
88	HTu	BTON, hyperpolarizability & per atom	-0.743	-0.551				
89	Bpa	B'M, No. bonds per atom: B is No. bonds	-0.165	0.633				
90	L	No. non-II atoms along the longest bond chain	0.215	0.540				
93	Xmin	minimum X coordinate	-0.398	-0.528				
97 98	Ymux	maximum Y coordinate maximum Z coordinate	-0.638 -0.577	-0.138				
98	Zmax	macunum z. coorunare Ymas-Ymin, nuolecular box width	-0.543	-0.045				
	DZ.	Zanas-Zanin, molecular box triatri Zanas-Zanin, molecular box height	-0.538	-0.01.5				
	sighb	Nb/Sm, HB donors/acceptors surface density; Sm	0.496	0.533				
	-	is molecular outface area						
105	sighyd	Nh/Sm. hydropholsic curbons surface density	0.660	0.752				
106	sign	Na/Sm. aroustic carbons surface density	-0.347	0.686				
107	sigp	Np/Sm, polar atoms surface density	0.494	0.669				
108	sign sigon	Nr/Sm, ring atoms surface density ON/Sm, ON atoms surface density	-0.426 0.575	-0.671				
1109		NUB, non-H atoms per bond	0.375	0.595				
	Np2	(Np - 12F, square function of Np	-0.527	-0.787				
112	ON2	(ON - 10) ² , senarc function of ON	-0.463	-0.593				
113	wb/3	twh = 0.61 ² , square function of wh	-0.682	-0.784				
	wu2	two = 0.31 ² , square function of wa twp = 0.44 ² , square function of wp	-0.168	0.627				
115	wp2	twp - 0.41°, square function of wp	-0.682	-0.784				
116	Mrefn2	(Mrefn - 0.19), square function of Mrefn	-0.485	-0.745				
117	sigp2	tsig = 0.0221 ² ; sig = Nve/S0, valence electron surface density; Nve is No, valence electrons	-0.669	-0.674				
118	RD	ratio of actual and standard bond lengths sum-	0.232	0,662				
119	RD2	(RD - 0.98), square function of RD	0.583	0.002				
	RRD	1/RD, inverse of RD	-0.243	-0.673				
pMi and rCH	Conclusion coefficient with network participation coefficient with network participation of the network of the conclusion of the conclusi							
			SELECTED MOLECULAR DESCRIPTORS Table 3. Selected molecular descriptors for drugs 1 - 12					

PLS (PARTIAL LEAST SQUARES) AND PCR (PRINCIPAL COMPONENT REGRESSION) PREDICTION OF VmrA RESISTANCE



Prediction set of drugs and xenobiotics. *V. parahaemolyticus* is sensitive to clinically used drugs against this bacterium (13-22) and to some agents (23, 24). The bacterium is probably resistant to other xenobiotics from this set (25-30).

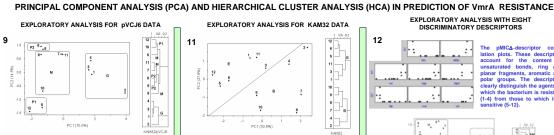
arameter	PLS (pVCJ6)	PCR (pVCJ6)	PLS (KAM)	PCR (KAM)
°Cs (%6)	L (70.0)	1 (70.0)	1 (52.8)	1 (53.6)
EV	0.721	0.683	0.452	0.443
FP	0.562	0.565	0.368	0.389
ł.	0.803	0.818	0.762	0.763
1	0.889	0.888	0.866	0.849
	0.410	0.410	0.288	0.270
F	0.206	0.209		
Ĵh			-0.228	-0.260
WT .	-0.204	-0.212		
9			-0.231	-0.261
IOMO			0.223	0.190
ighyd	-0.215	-0.217	-0.250	-0.258
ф2	-0.225	-0.215		
vh2			-0.258	-0.250
Sec. (m)	0.213	0.210		

Metric - -0.213 - -0.216 Regression vertices and advisical parameters are given for 72.5 and PCR modelling of efflux attrity of the 1- coli strain AAM22 (AVA) or (AAM25 - (AAA) the tarty coring and the tarty of advisors and the tarty correspondence of the tarty coring ATV- - underd error or disalides, ET - - underd error or prediction; Q - currelation secification of ulidation, R - - currelation coefficient of prediction; A - average absolute division for predicted trace prediction; A - average absolute division for predicted trace prediction; A - average absolute division of predicted trace prediction; A - average absolute division of predicted trace prediction; A - average absolute division of predicted trace prediction; A - average absolute division of predicted trace prediction; A - average absolute division of predicted trace prediction; A - average absolute division of predicted trace prediction; A - average absolute division; A - average absolute division

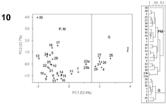
The PLS & PCR regression statistics for modeling of the both activities shows that there is no significant difference between the PLS and PCR models. The regression coefficients are in favour for pMIC(pVCJ6), whilst the errors are smaller for pMIC(rAM).

0.340 0.246 0.123 0.313 0.277 0.138 sensitive sensitive 1.059 1.052 1.123 1.117 resistant resistant resistant 23a 23b 0.872 0.968 resistant 24 0.844 0.810 resistant 0.196 0.230 25 1 774 1.872 1.506 resistant 0.523 0.499 26 27 1.455 (-) + 0.211 sensitive 12 0.247 1.604 1.671 resistant 28 29 30 12 0.411 0.392 consition 0.625 0.649 registant + 0.079 1.281 1.016 0.091 sensitive 1.302 resistant 15 0.087 sensitive 1.149 resistant (-)? 16 0.138 0.304 censitive The PLS & PCR regression models in predicting the MDR character of V. parahaemolyticus to 1-30. The cut-off criterion was 0.5 in

of V. parahaemolyticus to 1-30. The cut-off criterion was 0.5 in pMCa units, what seems not suitable for 3 gents. Other cut-off value or another methodology should be applied to eliminate wrong and doubtrill predictions. Error accumulation in both predicted pMIC(pVCJ6) and pMIC(KAM32) may contribute to wrong predictions. However, 20 of 31 (55%) agents were predicted correctly as agents to which the bacterium would be resistant or sensitive

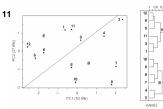


cores plot and complete dendogram with samples for the ining set. Clusters with good (6), moderately good (M) and or (P) substrates of VmrA are visible. *V. parahaemolyticus* is sistant to G and sensitive to M, P clusters. trainin

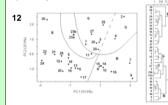


ores plot and complete dendogram with samples for the training+prediction set. According to the previous plots, it is clear that V. parahaemolyticus is sensitive to (13-24, 28, 30) and resistant to (25-27, 29).



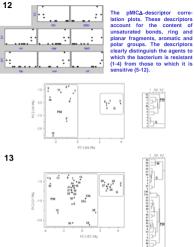


Scores plot and complete dendogram with samples for the training set. Clusters with more eloganted (E) and motionanched (B) molecules are visible, not related to the parahaemolyticus MDR character. dendogram with samples for the



Scores plot and complete dendogram with samples for the training-prediction set. The E-B discrimination is visible as in the previous plots. Beddes, the G (good WmA substrates) cluster occurs. This means that V. *paraheemolyticus* is resistant to 32-37 and 29, and semilive to other agents from the prediction set.

EXPLORATORY ANALYSIS WITH EIGHT DISCRIMINATORY DESCRIPTORS



Comparison of the exploratory analyses for the training (top) and clearly shows that the bacterium is sensitive to the PM and resistant to C clearly shows that the bacterium is sensitive to the PM and resistant to C cluster. This has been just observed in previous analyses.

eV

-13,003 -11,666 -11,251 -10,264 -6,345 -11,168 -11,168 -11,366 -13,862 -4,333 -6,123

163.649 215.097 203.297 241.318 129.924 175.499 356.612 219.420 316.371 249.845 176.495

0.4236 0.0094 0.4307 0.5931 0.8224 1.6859 0.6896 1.1533 2.3617 1.7979 1.5921 1.6083

0.8627 0.9600 0.8235 0.8333 0.3000 0.6957 0.6061 0.5625 0.5098 0.4250 0.6071

0.0500 0.0756 0.0100 0.0001 0.0176 0 0.153215 0.160000 0.170569 0.015625 0.037636 0.002601

0.0071

0.385641 0.385641 0.163210